Increased intron retention is linked to Alzheimer’s disease

Intron retention in aging and Alzheimer’s disease (AD): AD is an age-related neurodegenerative disorder with pathological accumulation of amyloid plaque (Masters et al., 2015), which can be classified into familial and sporadic form. In familial AD, mutations in genes encoding either amyloid precursor protein or presenilin (PS1 and PS2) cause overproduction of amyloid–β molecules and early onset of dementia. Late-onset sporadic AD, which accounts for majority of the cases (> 95%), is characterized by high degree of genetic and pathological heterogeneity. Although aging and genetic variants are two risks factors for sporadic AD, it remains unclear how epigenetic alterations during aging may contribute to its etiology. To determine the transcriptional changes that are associated with aging, we analyzed the transcriptome of head or brain tissues isolated at different ages from adult Drosophila, mice and human (Adusumalli et al., 2019). We observed an increase in the level of intron retention (IR) in the mRNA transcripts during aging across different species. These retained introns share highly conserved features and surprisingly, do not affect the expression level of their mRNA transcripts. As many age-dependent IR mRNA transcripts overlapped with curated AD genes, we further compared the IR patterns in the cerebral cortex and cerebellum isolated from large AD cohorts with their age-matched controls. Consistent with reports of aberrant splicing in AD (Tollervey et al., 2011; Bai et al., 2013; Raj et al., 2018), we found a significant increase in the rate of IR at genes that are involved in RNA processing and protein homeostasis in AD samples. Furthermore, many of these IR genes showed significant changes in their protein levels when compared to age-matched controls. Taken together, our findings suggest that increased IR is a transcriptional signature that is conserved across species during aging and may be linked to AD progression.

IR by alternative splicing has distinct biological roles in development and AD: Alternative splicing is a tightly regulated nuclear process that affects more than 95% of human genes and allows diversification of the proteome (Baralle and Giudice, 2017). Changes to the splicing patterns are prevalent during aging and its dysregulation has often been linked to neurodegenerative diseases like AD (Tollervey et al., 2011; Bai et al., 2013; Mazin et al., 2013; Raj et al., 2018). Alternative splicing can also cause specific intron(s) to be retained in the processed mRNA transcript. Indeed, many recent studies have shown that IR may play an active role in regulating gene expression and various biological processes via distinct mechanisms. In mammals, IR occurs at ~75% of genes that contained multiple exons (Braunschweig et al., 2014). The mRNA transcripts with retained introns are frequently degraded by nonsense-mediated mRNA decay (NMD) to remove those mRNAs whose expression is not required for the functioning of the specific cell types (Braunschweig et al., 2014). A similar mechanism is also in place to regulate the expression of functionally related genes during granulocyte differentiation (Wong et al., 2017). However, IR can also act as a strategy to allow rapid mobilization of selected mRNAs for protein translation in response to specific developmental cues. One example is the nuclear accumulation of specific mRNA transcripts with retained introns in mouse neocortical cells (Mauger et al., 2016). Upon N-methyl-D-aspartate receptor and calmodulin-dependent kinase mediated neuronal activation, these mRNA transcripts are spliced and exported to the cytosol for protein translation (Mauger et al., 2016). Therefore, IR eliminates the time needed for de novo transcription of large genes, allowing cells to respond to the inducing neuronal signals promptly. Similar to the results in cell lines and model organisms, genome-wide analyses with human cohorts also revealed widespread changes in the splicing patterns during aging and AD pathogenesis. Transcriptome analyses of newborns, young and old adult cerebral and prefrontal cortex revealed age-dependent changes in splicing patterns at about 40% of the expressed genes (Mazin et al., 2013). High-resolution microarray analysis of the middle temporal gyrus in the brain cortex region from healthy individuals and AD patients revealed significant differences in the exon usage at specific genes during aging and AD pathogenesis (Tollervey et al., 2011). Splicing abnormalities and intron retention were also reported in the frontal and dorsolateral prefrontal cortex of AD patients (Bai et al., 2013; Raj et al., 2018). While these studies highlighted the changes in splicing patterns during aging and AD pathogenesis, the characteristics of mRNA transcripts with IR are not fully explored. As such, we used IRFinder algorithm (Wong et al., 2017) to systematically evaluate the level of IR in the heads or brain tissues isolated from Drosophila, mouse and human at different ages, as well as three independent AD cohorts (Adusumalli et al., 2019). Compared to young adults, we observed increased IR of more than 340 genes in old flies. Surprisingly, the level of mRNA transcripts with retained introns remained unchanged, suggesting they may have evaded NMD. Genes with retained introns are enriched for distinct biological functions at different ages with approximately 15% of their human orthologues overlapped with curated AD genes from human DisGeNET database. In human prefrontal cortex, the differential IR genes are enriched for the processes that regulate proteostasis. This suggests that changes in IR patterns during aging can impact protein homeostasis, which may in turn contribute to AD aetiology.

Reports of aberrant splicing in AD brain tissues (Tollervey et al., 2011; Bai et al., 2013; Raj et al., 2018) suggest the likelihood of widespread changes in IR patterns and proteomic composition across different parts of the diseased brain. To test this possibility, we analyzed the transcriptome of cerebellum and frontal cortex from AD cohorts. Our analyses revealed significant differences in the levels of IR at many genes between AD and their age matched control tissues. In cerebellum, more than 3800 differential IR events were observed between age-matched control and AD tissues. Among these differential IR events, approximately 80% showed increased IR in AD cerebellum. Genes with differentially retained introns are enriched for pathways that regulate different aspects of mRNA and protein homeostasis. This suggests a feedback mechanism where changes in IR patterns may further enhance aberrant splicing and protein aggregation in AD cerebellum. Similarly, compared to healthy frontal cortex, there are more than 1100 increased IR events occurring at ~780 genes in AD tissues. IR does not lead to NMD as mRNA expression appeared unperturbed. However, ~15% of these increased IR genes show significant changes in their protein levels between healthy control and AD frontal cortex (Ping et al., 2018). These results support the notion that translation of mRNA transcripts with retained intron may contribute to the aberrant proteomic landscape in AD frontal cortex.

Although our study uncovered global alterations in IR patterns during aging and AD pathogenesis, the mechanisms underlying such changes remain unresolved. Recent evidence indicates that RNA splicing may be regulated through the interplay between genetic and epigenetic factors (Figure 1). Integrative analyses of transcriptomes and single nucleotide polymorphisms data from genome-wide association studies revealed that genetic variants at spliced acceptor/donor sites may impact the splicing or intron usage at some AD susceptibility loci (Raj et al., 2018). Retained introns were also shown to have higher CG content compared to the spliced introns, indicating that IR may be driven by the underlying DNA sequences (Adusumalli et al., 2019; Braunschweig et al., 2014; Wong et al., 2017). The strong correlation between RNA polymerase II stalling and higher level of retained introns at the CG-rich sequences on DNA suggests that IR may be regulated by the cross-talk between transcription and splicing machineries (Braunschweig et al., 2014). IR may also be regulated epigenetically given that CG-rich sequences are subjected to DNA methylation. Indeed, DNA methylation is required for the recruitment of MeCP2 and its interacting splicing factors to prevent IR during granulocytes differentiation (Wong et al., 2017). Insoluble aggregation of spliceosome component U1 small nuclear ribonucleoprotein A1 into tangles of the brain cortex region is also correlated with aberrant splicing in the frontal cortex (Bai et al., 2013). This suggests that protein aggregation of splicing machinery may be one of the underlying causes of increased IR in AD. Interestingly, it was recently reported that dietary restriction, which extended the lifespan of Caenorhabditis elegans, could reduce the level of IR during aging (Heintz et al., 2017). Taken together, these findings raise the intriguing
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